ORIGINAL ARTICLE

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Expression of bcl-2, Bax and Fas in oxyphil cells of Hashimoto thyroiditis

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Abstract Immunoreactivity for bcl-2, Bax and Fas was analysed in 16 cases with Hashimoto thyroiditis. Bcl-2expression was constantly seen in regular thyrocytes and in the mantle-zone of lymphofollicular infiltrates. However, thyrocytes in the vicinity of lymphoid infiltrates and, especially, mitochondria-rich oxyphil cells exhibited reduced staining or none at all for bcl-2. Bax was found to be weakly reactive or negative in normal thyrocytes and was not up-regulated in bcl-2-deficient epithelial cells. In contrast, expression of Fas was markedly increased both in typical thyrocytes and in oxyphil cells within areas of lymphocytic infiltration. In conclusion, focal lack of bcl-2 expression together with up-regulation of Fas is a constant feature of Hashimoto thyroiditis. The reaction pattern of oxyphil cells is identical to that of affected typical thyrocytes without proliferation of mitochondria. Loss of bcl-2 with up-regulation of Fas is therefore likely to precede oncocytic change. Whether these alterations are involved in the process of oncocytic transformation remains to be clarified, however.

Key words Thyroid \cdot Hashimoto thyroiditis \cdot Oxyphil cell \cdot Immunohistochemistry \cdot bcl-2 \cdot Bax \cdot Fas

Introduction

Oxyphil cell metaplasia of the thyroid is a characteristic feature of Hashimoto thyroiditis [15, 42, 43]. The significance of this cell type is still unclear, however. In recent studies we have shown that defects of cytochrome-coxidase (complex IV of the respiratory chain) and a 4977-bp deletion (common deletion) of mitochondrial DNA may occur in the cells, which are very rich in mitochondria [38]. Furthermore, glucose transporter type IV is highly expressed in these cells [39].

The bcl-2 gene was first described in follicular lymphomas in which the chromosomal translocation, t (14; 18) juxtaposes the bcl-2 gene to the immunoglobulin gene locus, resulting in high bcl-2 protein expression and prevention of apoptosis [58, 59]. As well as being expressed in various tumors, including carcinomas of the thyroid [1, 4, 8, 11, 19, 30, 41, 46–49, 53–55], bcl-2 is usually expressed in nonneoplastic lymphocytes and in regular tissue of the thyroid [30, 48, 49], the breast [25], the endometrium [21, 61] and the adrenals [9].

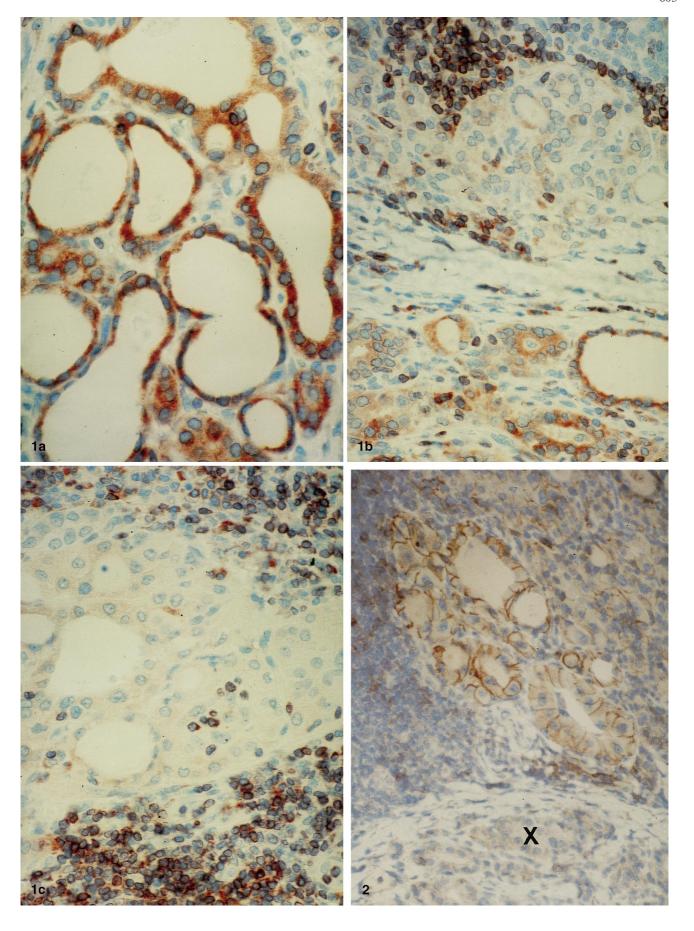
Bcl-2 expression in the thyroid has been found to be associated with differentiated follicular and papillary carcinomas, whereas anaplastic carcinomas, but also oncocytic tumors, mostly lack bcl-2 expression [30, 37, 48, 49]

In contrast to bcl-2, which prevents cell death, Bax protein promotes cell death by forming homo- and heterodimers with bcl-2 [45].

In thyroid carcinomas Bax protein has been found to be almost completely lacking in well-differentiated carcinomas and to be coexpressed with bcl-2 in poorly differentiated carcinomas, but it is selectively present in anaplastic carcinomas [30].

In recent studies Apo-1/Fas, a member of the TNF receptor family involved in triggering apoptosis [44, 64], has been shown to be up-regulated in Hashimoto thyroiditis [10, 14, 32]. The reactivity of oxyphil cells has not, however, been analyzed specifically.

In the present study we investigated the reaction pattern of oxyphil cells for Fas, Bax, bcl-2 and were especially interested to find whether oxyphil cells in Hashimoto thyroiditis lack bcl-2 expression, as is the case in oncocytic neoplasms of the thyroid gland [37]. Furthermore, the reactivity of oxyphil cells was compared with the reaction profile of affected nonoxyphil thyrocytes to see whether alterations of these proteins might be involved in the pathogenesis of oxyphil metaplasia in Hashimoto thyroiditis.



Materials and methods

Sixteen resection specimens from patients with a diagnosis of Hashimoto thyroiditis were obtained from the files of the Department of Pathology of the University of Munich for immunohistochemical analysis. The male-to-female ratio was 1:3, and the median age of the patients, 51 years (mean age 54 years, range 27–77 years).

Surgical resection had been performed mainly because of goitrous enlargement partly caused by the additional presence of endemic goitrous nodules and because of unclear cytological results of fine-needle biopsies.

The diagnosis of Hashimoto thyroiditis was made mainly with reference to morphological criteria. These encluded a bilateral involvement, the presence of a predominantly diffuse lymphocytic infiltrate, often with formation of germinal centers, atrophy and destruction of follicular epithelium associated with oxyphilic cell metaplasia.

Immunohistochemical analysis was performed on formalinfixed paraffin-embedded 5-μm sections, deparaffinized and rehydrated through graded alcohols. Immunocytochemical reactivity was tested for bcl-2 (monoclonal, mouse, 1:10, Dako), Bax (polyclonal, rabbit, 1:20, Calbiochem) and Fas (monoclonal, mouse, 1:40, Dako).

The slides were subjected to pretreatment by microwaving in citrate cytobuffer at pH 6 in a 750-W oven for 2×10 min. Before further staining slides were also pretreated with avidin/biotin blocking reagent (Dako) for 15 min each at room temperature.

The specific antibodies were applied for 30 min at room temperature. Detection was performed with streptavidin-conjugated peroxidase (Dako) and for Bax protein with ABC Elitekit (Vector), using aminoethylcarbazole (AEC) as chromogen.

For negative controls the primary antibody was omitted and the primary antibody was substituted by nonimmune immunoglobulin of the same class as the primary antibody.

Results

Routine hematoxylin and eosin staining disclosed the typical features of Hashimoto thyroiditis with a prominent diffuse and lymphofollicular infiltrate irregularly distributed in the thyroid tissue and multiple foci of follicular epithelium destruction. To a varying extent pronounced oxyphil change of thyrocytes was seen (Fig. 1C).

Bcl-2

Bcl-2 staining showed a cytoplasmic type of reaction in normal thyrocytes distant from lymphocytic infiltrates (Fig. 1A), which was identical to the staining patterns in

◆ Fig. 1A-C Bcl-2-immunohistochemistry. A Normal thyroid follicles with cytoplasmic reactivity. B Nonoxyphilic thyroid follicles in the vicinity of lymphocytic infiltrates are devoid of immunohistochemically detectable bcl-2 in the cytoplasm. C In oxyphil thyrocytes the typical cytoplasmic reaction is also lacking. B, C Lymphocytes stain positive. ×540

Fig. 2 Immunohistochemical detection of Fas. In areas with inflammatory infiltrates intensive expression of Fas is seen with a predominantly membranous staining pattern in oxyphil cells. Adjacent regular thyroid follicles (X) show only weak expression or none at all. $\times 350$

thyroids not affected by inflammatory disease. The reaction was equally developed in all follicles, and a high staining intensity predominated. Typically the lymphocytic infiltrates also were stained (Fig. 1B, C), the germinal centers being spared. In contrast, expression of bcl-2 was reduced or lacking in small aggregates of nonoxyphilic thyrocytes closely associated with lymphocytic infiltrates (Fig. 1B). Oxyphil cells in particular were predominantly negative for bcl-2 (Fig. 1C). Only occasionally was a faint cytoplasmic reaction seen (not shown).

Bcl-2 staining was lacking both in single oxyphil cells and in oxyphilic cell clusters with and without inflammatory infiltrate in its vicinity.

Bax

The antibody to Bax was found to be weakly reactive or negative in most regular thyrocytes and oxyphil cells, whereas immunoreactivity was seen especially in the germinal center cells of lymphofollicular infiltrates (not shown).

Fas

Thyrocytes from normal thyroid tissue showed only a weak reaction or none at all in the cytoplasm. A membranous staining pattern was not seen. In contrast, thyroid follicles near or within lymphocytic infiltrates exhibited intensive staining (Fig. 2). The positive staining reaction was seen both in nonoxyphil follicular cells and in typical oxyphil cells. Especially in oxyphil cells the staining pattern was mainly membranous. A clear positive reaction was also observed in the infiltrating lymphocytes, particularly at the center of lymphoid follicles, whereas the reaction was predominantly lacking in the adjacent mantle zone.

Discussion

Bcl-2 belongs to a group of genes (bcl-2 gene family) that modulates cell death [3]. The expression of the gene family seems to be regulated differentially among cell types and stage of differentiation. The biological effect on a cell is therefore dependent on its level, selective expression and dimerization status [28]. Bcl-2 is expressed in several normal and various malignant human tissues [17, 27, 28]. Interestingly, bcl-2 is expressed in most developing epithelial tissues and appears to be restricted to cells at specific stages of differentiation [27, 28].

In adult epithelia bcl-2 is found in duct cells of exocrine cells, including pancreas, salivary and sweat glands [28], and in hormone-responsive cells, such as those in the endometrium, mammary glands, prostate, adrenals [9, 21, 25, 61] and thyroid [30, 48, 49].

In thyroid carcinomas bcl-2 expression depends on the degree of differentiation, being absent in anaplastic tumors [30, 48, 49]. This finding is in agreement with the idea that bcl-2 expression allows epithelia to differentiate and morphogenesis to proceed [28].

Interestingly, anaplastic thyroid carcinomas express the Bax protein exclusively, which forms homo- and heterodimers with bcl-2 in vivo [30]. Thereby, bcl-2 is inactivated and programmed cell death may proceed [45].

We analyzed the expression of bcl-2, Bax and Fas in Hashimoto thyroiditis with special reference to the reaction pattern of the mitochondria-rich oxyphil cells. It was found that bcl-2 is down-regulated in Hashimoto cells, particularly at inflammatory sites, down-regulation being most evident in oxyphil cells. Conversely, Fas was found to be up-regulated in thyrocytes near the inflammatory reaction sites, with oxyphil cells showing an intensive, often membranous staining reaction.

The data on expression of bcl-2 and Fas have been corroborated by two recent studies [14, 32]. In these studies the reactivity of mitochondria-rich oxyphil cells was not specifically analyzed, however. Furthermore, our results indicate that up-regulation of Bax is not apparently involved in the pathogenesis of Hashimoto thyroiditis. This is consistent with the finding that apoptosis in Hashimoto thyroiditis is induced by Fas-ligand up-regulation [10, 14, 32]. Since Fas belongs to the TNF receptor family, it is interesting to note that TNF receptor II has recently been localized in the inner mitochondrial membrane of preadipocytes [24].

Bcl-2 has been localized to the outer membrane of mitochondria, in a patchy distribution probably related to contact sites between the outer and inner mitochondrial membrane [22, 34, 52]. A variety of mitochondrial events such as the sensitivity to oligomycin inhibition of complex V, to cyanide and the release of sequestrated calcium in the matrix appear to be modulated by bcl-2 [12]. Disruption of the transmembrane potential (electrochemical proton gradient) of the inner mitochondrial membrane is an early event in apoptosis, which can be prevented by bcl-2 overexpression [23]. Bcl-2 blocks apoptosis in cells without mitochondrial DNA and therefore with a nonfunctional respiratory chain [18, 31].

What precisely causes dysregulation of bcl-2 and Fas genes in Hashimoto thyroiditis is not yet known. Downregulation of bcl-2 has been shown to be the first step in Fas-mediated apoptosis in the male reproductive tract [57], and thyroid-stimulating hormone (TSH) inhibits Fas-mediated apoptosis [20]. The altered immunohistochemical profile of bcl-2 and Fas, especially in the vicinity of inflammatory infiltrates, is most consistent with the involvement of cytokine effects. In fact, TNF alpha has been shown to decrease bcl-2 expression and upregulate Fas in human cell lines [5, 29]. In addition, interleukin-1 beta has been shown to up-regulate Fas and induce apoptosis in Hashimoto thyroiditis [10]. Recently a deletion of C-terminal signal anchor sequence has been found to interfere with targeting of bcl-2 to the outer mitochondrial membrane and with bcl-2-mediated suppression of apoptosis [23]. Furthermore, it should be mentioned that bcl-2 is down-regulated by the suppressor gene *p53* [13, 33].

We recently reported on reduced expression of bcl-2 in oncocytic adenomas and carcinomas of the thyroid [37]. In these tumors no up-regulation of Fas could be found, however (unpublished results). Since the lack of bcl-2 in anaplastic thyroid carcinomas [30, 48, 49] is associated with up-regulation of Bax – which was not the case in the bcl-2-negative oxyphil cells in Hashimoto thyroiditis – it may be assumed that pathogenetic mechanisms are different in neoplastic and metaplastic oxyphilic transformation.

The pathogenesis of oncocytic transformation, and thus of mitochondrial proliferation, is still unresolved. Generally the rate of ATP consumption, hypoxia and nuclear-regulatory factors are assumed to be of importance for the regulation of mitochondrial proliferation. The role of cytokines is still unclear. Our results have shown that the immunoreactive profile of oxyphil cells for bcl-2, Bax, and Fas does not differ from affected follicular cells without mitochondrial proliferation. Dysregulation of bcl-2 and Fas is therefore likely to precede oncocytic metaplasia and could be involved in its pathogenesis. Mitochondrial DNA, at least, is not a necessary prerequisite, since mitochondrial proliferation is also seen in case of depletion of mitochondrial DNA occurring under various conditions [2, 6, 26, 35, 36, 60].

Various studies indicate that mitochondria are possible primary sites for the action of thyroid hormones, and especially of T3 [7, 16, 50, 51, 56, 62, 63], and it has been shown that T3 induces biogenesis of mitochondria [50, 51]. Furthermore, a 43-kDa protein related to c-ErbA alpha 1 has been located in the mitochondrial matrix and could act as a mitochondrial T3-dependent transcription factor in a similar way to its function in the nucleus [63]. It would be interesting to know whether there is a change in the receptor status for TSH and thyroid hormones in oxyphilic cells.

In summary, we have shown that focal loss of bcl-2 and up-regulation of Fas is a characteristic feature of Hashimoto thyroiditis occurring prior to oncocytic metaplasia. It remains to be seen, however, whether the altered cells without a typical oxyphil appearance are already harboring some type of nuclear and/or mitochondrial genetic alteration responsible for mitochondrial proliferation. These factors are still unclear.

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References

- Baretton GB, Diebold J, Christotoris G, et al (1996) Apoptosis and immunohistochemical bcl-2 expression in colorectal adenomas and carcinomas. Cancer 77:255–264
- Bertini E, Vici CD, Servidei S, et al (1992) Fatal infantile liver failure associated with mitochondrial DNA depletion. J Pediatr 121:896–901

- Boise LH, Gonzales-Garcia M, Postema CE et al. (1993) bcl-x, a bcl-2-related gene that functions as a dominant regulator of apoptotic cell death. Cell 74:597–608
- Bubendorf L, Sauter G, Moch H, et al (1996) Prognostic significance of bcl-2 in clinically localized prostate cancer. Am J Pathol 148:1557–1565
- Chen M, Quintans J, Fuks Z, et al (1995) Suppression of Bcl-2 messenger RNA production may mediate apoptosis after ionizing radiation, tumor necrosis factor alpha and ceramide. Cancer Res 55:991–994
- Dalacas MC, Pezeshkpour I, et al (1990) Mitochondria myopathy caused by longterm zidovudine therapy. N Engl J Med 322:1098–1105
- 7. Demonacos C, Karayanni N, Hatzoglou E, et al (1996) Mitochondrial genes as sites of primary action of steroid hormones. Steroids 61:226–232
- 8. Diebold J, Baretton G, Felchner M, et al (1996) bcl-2-expression, p53 accumulation, and apoptosis in ovarian carcinomas. Am J Clin Pathol 105:341–349
- Fogt F, Vortmeyer AO, Poremba C, et al (1998) Bcl-2 expression in normal adrenal glands and in adrenal neoplasms. Mod Pathol 11:716–720
- Giordano C, Stassi G, De Maria R, et al (1997) Potential involvement of Fas and its ligand in the pathogenesis of Hashimoto's thyroiditis. Science 275:960–963
- 11. Gorczyca W, Markiewski M, Kram A, et al (1995) Immunohistochemical analysis of bcl-2 and p53 expression in breast carcinomas: their correlation with Ki-67 growth fraction. Virchows Arch 426:229–233
- 12. Green DR, Reed JC, et al (1998) Mitochondria and apoptosis. Science 281:1309–1312
- Haldar S, Negrini M, Monne M, et al (1994) Down-regulation of bcl-2 by p53 in breast cancer cells. Cancer Res 54:2095– 2097
- 14. Hammond LJ, Lowdell MW, Cerrano PG, et al (1997) Analysis of apoptosis in relation to tissue destruction associated with Hashimoto's autoimmune thyroiditis. J Pathol 182:138–144
- Hashimoto H (1962) Zur Kenntnis der lymphomatösen Veränderungen der Schilddrüse (Struma lymphomatosa). Arch Klin Chir 97:219–248
- Hashizume K, Ichikawa K (1982) Localization of 3,5,3'-L-triiodothyronine receptor in rat kidney mitochondrial membranes. Biochem Biophys Res Commun 106:920–926
- Hockenbery DM, Zutter M, Hickey W, et al (1991) Bcl-2 is topographically restricted in tissues characterized by apoptotic cell death. Proc Natl Acad Sci USA 88:6961–6965
- Jacobson MD, Burne JF, King MP, et al (1993) Bcl-2 blocks apoptosis in cells lacking mitochondrial DNA. Nature 361: 365–369
- Joensuu H, Phylkkänen L, Toikkanen S (1994) bcl-2-protein expression and long-term survival in breast cancer. Am J Pathol 145:1191–1198
- Kawakami A, Eguchi K, Matsuoka N, et al (1996) Thyroid-stimulating hormone inhibits Fas antigen-mediated apoptosis of human thyrocytes in vitro. Endocrinology 137:3163–3169
- 21. Koh EA, Illingworth PJ, Duncan WC, et al (1995) Immunolocalization of bcl-2 protein in human endometrium in the menstrual cycle and simulated early pregnancy. Hum Reprod 10: 1557–1562
- 22. Krajewski S, Tanaka S, Takayama S, et al (1993) Investigation of the subcellular distribution of the bcl-2 oncoprotein: residence in the nuclear envelope, endoplasmic reticulum, and outer mitochondrial membranes. Cancer Res 53:4701–4714
- Kroemer G, Zamzami N, Susin SA (1997) Mitochondrial control of apoptosis. Immunol Today 18:44–51
- Ledgerwood EC, Prins JB, Bright NA, et al (1998) Tumor necrosic factor is delivered to mitochondria where a tumor necrosis factor-binding protein is localized. Lab Invest 78:1583–1589
- 25. Leek RD, Kaklamanis L, Pezella F, et al (1994) bcl-2 in normal human breast and carcinoma, association with oestrogen receptor-positive, epidermal growth factor receptor-negative tumours and in situ cancer. Br J Cancer 69:135–139

- Lewis W, Dalacas MC (1995) Mitochondrial toxicity of antiviral drugs. Nat Med 1:417–422
- Lu QL, Poulson R, Wong L, et al (1993) Bcl-2 expression in adult and embryonic non-haematopoietic tissues. J Pathol 169:431–437
- 28. Lu QL, Abel P, Foster CS, et al (1996) Bcl-2 role in epithelial differentiation and oncogenesis. Hum Pathol 27:102–110
- 29. Maciejewski J, Selleri C, Anderson S, et al (1995) Fas antigen expression on CD 34+ human marrow cells is induced by interferon gamma and tumor necrosis factor alpha and potentiates cytokine mediated hematopoietic supression in vitro. Blood 85:3183–3190
- Manetto V, Lorenzini R, Cordon-Cardo C, et al (1997) Bcl-2 and bax expression in thyroid tumours. An immunohistochemical and Western blot analysis. Virchows Arch 430:125–130
- Marchetti Ph, Susin SA, Decaudin D, et al (1996) Apoptosisassociated derangement of mitochondrial function in cells lacking mitochondrial DNA. Cancer Res 56:2033–2038
- 32. Mitsiades N, Poulaki V, Kotoula V, et al (1998) Fas/Fas ligand up-regulation and Bcl-2 down regulation may be significant in the pathogenesis of Hashimoto's thyroiditis. J Clin Endocrinol Metab 83:2199–2203
- 33. Miyashita T, Krajewski S, Krajewska M, et al (1994) Tumor supressor p53 is a regulator of bcl-2 and bax gene expression in vitro and in vivo. Oncogene 9:1799–1805
- Monaghan P, Robertson D, Amos TAS, et al (1993) Ultrastructural localization of bcl-2 Protein. J Histochem Cytochem 40:1819–1825
- Moraes CT, Shanske S, Tritschler HJ, et al (1991) MtDNA depletion with variable tissue expression: a novel abnormality in mitochondrial disease. Am J Hum Genet 48:492–501
- Morais R, Zinkewich-Péotti K, Parent M, et al (1994) Tumorforming ability in athymic nude of human cell lines devoid of mitochondrial DNA. Cancer Res 54:3889–3896
- 37. Müller-Höcker J (1999) Immunoreactivity of p53, Ki 67 and Bcl2 in oncocytic adenomas and carcinomas of the thyroid gland. Hum Pathol 30:926–933
- 38. Müller-Höcker J, Jacob U, Seibel P (1998) Hashimoto thyroiditis is associated with defects of cytochrome-c oxidase in oxyphil Askanazy cells and with the common deletion (4.977) of mitochondrial DNA. Ultrastruct Pathol 22:91–100
- Müller-Höcker J, Schäfer A, Strowitzki T (1998) Glucose transporter 4 (Glut 4) is highly expressed in mitochondria rich oxyphil cells. Appl Immunohistochem 6:224–227
- Nakamura S, Akazawa K, Kinukawa N, et al (1996) Inverse correlation between the expression of bcl-2 and p53 proteins in primary gastric lymphoma. Hum Pathol 27:225–233
- in primary gastric lymphoma. Hum Pathol 27:225–233 41. Nakopoulou L, Vourlakou C, Zervas A, et al (1996) The prevalence of bcl-2, p53, and Ki-67 immunoreactivity in transitional cell bladder carcinomas and their clinicopathologic correlates. Hum Pathol 29:146–154
- 42. Nesland JM, Sobrinho-Simoes MA, Holm R, et al (1985) Hürthle-cell lesions of the thyroid: a combined study using transmission electron microscopy, scanning electron microscopy, and immunocytochemistry. Ultrastruct Pathol 8:269–290
- 43. Nève P (1969) The ultrastructure of thyroid in chronic autoimmune thyroiditis. Virchows Arch [A] 346:302–317
- 44. Oehm A, Berhmann I, Falk W, et al (1992) Purification and molecular cloning of the APO-1 cell surface antigen, a member of the TNF/NGF receptor superfamily. J Biol Chem 267:10709–10715
- 45. Oltvai Z, Milliman C, Korsmeyer SJ (1993) Bcl-2 heterodimerizes in vivo with a conserved homolog, Bax, that accelerates programmed cell death. Cell 74:609–619
- Pezella F, Turley H, Kuzu I, et al (1993) Bcl-2 protein in nonsmall-cell lung carcinoma. N Engl J Med 329:690–694
- 47. Pezella F, Morrison H, Jones M, et al (1993) Immunohistochemical detection of p53 and bcl-2 proteins in non-Hodgkin's lymphoma. Histopathology 22:39–44
- 48. Pilotti S, Collini P, del Bo R, et al (1994) A novel panel of antibodies that segregates immunocytochemically poorly differentiated carcinoma from undifferentiated carcinoma of the thyroid gland. Am J Surg Pathol 18:1054–1064

- 49. Pilotti S, Collini P, Rilke F, et al (1994) Bcl-2 protein expression in carcinomas originating from the follicular epithelium of the thyroid gland. J Pathol 172:337–342
- Reith A (1973) The influence of triiodothyronine and riboflavin deficiency on the rat liver with special reference to mitochondria. Lab Invest 29:216–228
- Reith A, Brdiczka D, Nolte J, et al (1973) The inner membrane of mitochondria under influence of triiodothyronine and riboflavin deficiency in rat heart muscle and liver. Exp Cell Res 77:1–14
- 52. Riparbelli MG, Callaini G, Tripodi SA, et al (1995) Localization of the bcl-2 protein to the outer mitochondrial membrane by electron microscopy. Exp Cell Res 221:363–369
- 53. Saegusa M, Takano Ý, Hashimura M, et al (1995) The possible role of bcl-2 expression in the progression of tumors of the uterine cervix. Cancer 76:2297–2303
- Saegusa M, Takano Y, Okayasu I (1995) Bcl-2 expression and its association with cell kinetics in human gastric carcinomas and intestinal metaplasia. J Cancer Res Clin Oncol 121: 357–363
- Silvestrini R, Veneroni S, Daidone MG, et al (1994) The BCl-2-protein: a prognostic indicator strongly related to p53 protein in lymph node-negative breast cancer patients. J Natl Cancer Inst 86:499–504
- 56. Sterling K, Milch PO, Brenner MA, et al (1977) Thyroid hormone action: the mitochondrial pathway. Science 197:996–

- 57. Suzuki A, Matsuzawa A, Iguchi T (1996) Down-regulation of Bcl-2 is the first step on Fas-mediated apoptosis of male reproductive tract. Oncogene 13:31–37
- Tsujimoto Y, Cossmann J, Jaffe E, et al (1985) Involvement of the bcl-2 gene in human follicular lymphoma. Science 228: 1440–1443
- Tsujimoto Y, Croce CM (1986) Analysis of the structure, transcripts, and protein products of bcl-2, the gene involved in human follicular lymphoma. Proc Natl Acad Sci USA 83: 5214–5218
- Vormier V, Rötig A, Quartino AR et al. (1990) Widespread multitissue deletions of the mitochondrial genome in the Pearson marrow-pancreas syndrome. Clin Lab Obstet 117: 599–602
- 61. Watanabe H, Kanzaki H, Narukawa S, et al (1997) bcl-2 and Fas expression in eutopic and ectopic human endometrium during the menstrual cycle in relation to endometrial cell apoptosis. Am J Obstet Gynecol 176:360–368
- Wrutniak C, Cabello G (1996) La voie d'action mitochondriale directe de la triiodothyronine mythe ou réalité? Med Sci 12:475–484
- 63. Wrutniak C, Cassar-Malet I, Marschal S, et al (1995) A 43-kDa protein related to c-Erb A alpha 1 is located in the mitochondrial matrix of rat liver. J Biol Chemistry 270: 16347–16357
- 64. Wyllie AM (1995) The genetic regulation of apoptosis. Curr Opin Genet Dev 5:97–104